

101.201-48/12 – including *Taq* pol., IFU-01  
 101.201-48u/12u – without *Taq* pol., IFU-02

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 “Instructions for Use” (IFU)

Lot No.: **4E0**

Lot-specific information

**Olerup SSP® DQ low resolution**

<b>Product number:</b>	101.201-48/12 – including <i>Taq</i> pol. 101.201-48u/12u – without <i>Taq</i> pol.
<b>Lot number:</b>	4E0
<b>Expiry date:</b>	2019-04-01
<b>Number of tests:</b>	48 tests – Product No. 101.201-48/48u 12 tests – Product No. 101.201-12/12u
<b>Number of wells per test:</b>	15+1
<b>Storage - pre-aliquoted primers:</b>	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

**This Product Description is only valid for Lot No. 4E0.**

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP®  
 DQ LOW RESOLUTION LOT (92Y)**

One well has been added to DQ low resolution, well **16**.

The DQ low resolution specificity and interpretation tables have been updated for the HLA-DQB1 alleles described since the previous *Olerup SSP®* DQ low resolution lot was made (**Lot No 92Y**). The kit design is based on IMGT/HLA database 3.25.0.

As of lot series V, the Specificity Table is included in the lot-specific Product Insert, and the Interpretation Table is included in the Worksheet.

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The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
1	Added	-	5'-primer added for the DQB1*05:105 allele.
2	-	Added, Modified, Removed	3'-primer added for the DQB1*06:199 allele, 3'-primer modified for improved HLA-specific amplification. 3'-primers removed.
3	Added	-	5'-primers added for the DQB1*02*40 and DQB1*02:72 alleles.
4	-	Added	3'-primer added for the DQB1*03:154 allele. 5' and 3'-primer have been optimized for increased yield.
8	Added	Added	5'-primer added for the DQB1*04:30 allele, 3'-primer added for the DQB1*04:02:08 allele.
9	Modified	-	5'-primer modified for increased yield of the DQB1*04:01 alleles.
10	-	Added	3'-primer added for the DQB1*05:98 allele.
15	Added	Added	Negative Control moved to well 16, Primer pair added for the allelic resolution of the DQB1*03:10 alleles.
16	-	-	Negative Control added from well 15.

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Well **16** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP*® HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.

The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80	85
<b>5'-primer<sup>1</sup></b>	<b>164</b>	<b>340</b>	<b>440</b>	<b>45</b>	<b>45</b>	<b>43</b>	<b>36</b>
	5'-CAC <sup>3'</sup>	5'-Agg <sup>3'</sup>	5'-TTA <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-TAC <sup>3'</sup>
							5'-TAT <sup>3'</sup>
<b>3'-primer<sup>2</sup></b>	<b>231</b>	<b>2<sup>nd</sup> I</b>	<b>507</b>	<b>59</b>	<b>58</b>	<b>57</b>	<b>47</b>
	5'-TgC <sup>3'</sup>	5'-AAA <sup>3'</sup>	5'-TTg <sup>3'</sup>	5'-CTC <sup>3'</sup>	5'-ggC <sup>3'</sup>	5'-CTC <sup>3'</sup>	5'-ACA <sup>3'</sup>
							5'-gCA <sup>3'</sup>
							5'-gCC <sup>3'</sup>
							5'-TgT <sup>3'</sup>
<b>A*</b>	<b>+</b>	<b>+</b>	<b>+</b>				
<b>B*</b>	<b>+</b>	<b>+</b>	<b>+</b>				
<b>C*</b>	<b>+</b>	<b>+</b>	<b>+</b>				
<b>DRB1</b>				<b>+</b>	<b>+</b>		
<b>DRB3</b>				<b>+</b>	<b>+</b>		
<b>DRB5</b>				<b>+</b>			
<b>DQB1</b>					<b>+</b>		
<b>DPB1</b>						<b>+</b>	
<b>DQA1</b>							<b>+</b>

<sup>1</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>2</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon or the 2<sup>nd</sup> intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

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## PRODUCT DESCRIPTION

### DQ low resolution SSP typing

#### CONTENT

The primer set contains 5'- and 3'-primers for grouping the DQB1 alleles into the serological groups DQ2 to DQ9.

*Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.*

#### PLATE LAYOUT

Each test consists of 16 PCR reactions in a 16 well PCR plate.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	NC

The 16 well cut PCR plate is marked with 'DQ LOW' in silver/gray ink.

Well No. 1 is marked with the Lot No. '4E0'.

Wells 1 to 15 – DQ low resolution primers.

Well 16 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are covered with a PCR-compatible foil.

**Please note:** When removing each 16 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

#### INTERPRETATION

Only the DQB1 alleles will be amplified by the DQ low resolution typing kit. Thus, the interpretation of DQ low resolution typings is not influenced the DQB2 and DQB3 genes.

#### UNIQUELY IDENTIFIED ALLELES

All the DQB1 alleles, i.e. **DQB1\*05:01:01:01 to 05:123**, **DQB1\*06:01:01 to 06:209**, **DQB1\*02:01:01 to 02:74**, **DQB1\*03:01:01:01 to 03:230** and **DQB1\*04\*01:01 to 04:36N**, recognized by the HLA Nomenclature Committee in July 2016<sup>1,2</sup> will be amplified by the primers in the DQ low resolution SSP kit. The DQB1 alleles will be grouped into their corresponding serological specificities, i.e.:

DQ5(1) =	DQB1*05:01-05:05 <sup>3</sup>
DQ6(1) =	DQB1*06:01-06:33 <sup>3</sup>
DQ2 =	DQB1*02:01-02:05
DQ3 =	DQB1*03:01-03:20 <sup>3</sup>
DQ4 =	DQB1*04:01-04:02 <sup>3</sup>

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The DQ3 alleles may be further subdivided into the DQ3, DQ7, DQ8 and DQ9 based upon serology and expert assignment. Thus;

DQ3 = DQB1\*03:06, 03:10, 03:14  
DQ7 = DQB1\*03:01:01-03:01:03, 03:04, 03:09, 03:13, 03:16, 03:19  
DQ8 = DQB1\*03:02:01, 03:05:01, 03:07, 03:08, 03:11, 03:18  
DQ9 = DQB1\*03:03:02, 03:12, 03:15, 03:17, 03:20

<sup>1</sup>HLA-DQB1 alleles listed on the IMGT/HLA web page 2016-July-14, release 3.25.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

<sup>2</sup>Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

<sup>3</sup>The serological split of the DQB1\*05:05 to 05:123, DQB1\*06:06 to 06:07, 06:10, 06:13, 06:15 to 06:24 and 06:27 to 06:209, the DQB1\*02:04 to 02:74 the DQB1\*03:02:02 to 03:02:04, 03:03:03, 03:05:02, 03:07 to 03:09 and 03:11 to 03:230 and the DQB1\*04:03:01 to 04:36N alleles is not known. The grouping of not serologically defined alleles is taken from the expert-assigned serological grouping in Tissue Antigens (2009) 73:95-170.

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Lot-specific information  
**SPECIFICITY TABLE**

**DQ low resolution SSP typing**

Specificities and sizes of the PCR products of the 15+1 primer mixes used for DQ low resolution SSP typing

Primer Mix	Size of spec. PCR product <sup>1</sup>	Size of control band <sup>2</sup>	DQ serology <sup>3</sup>	Amplified DQB1 alleles <sup>4</sup>
1 <sup>7</sup>	135 bp, 230 bp	<b>515 bp</b>	5	*05:01:01:01-05:01:15, 05:01:17-05:59, 05:61-05:81, 05:83-05:97, 05:99-05:115, 05:117-05:123
2 <sup>7</sup>	135 bp, 185 bp, 220 bp, 270 bp	<b>515 bp</b>	1, 5, 6	*03:23:01-03:23:02, 03:217, 04:10, 06:01:01-06:168, 06:170-06:209
3	210 bp	430 bp	2	*02:01:01-02:74
4	130 bp, 220 bp	<b>515 bp</b>	3, 7	*03:01:01:01-03:01:33, 03:04:01-03:04:02, 03:09-03:10:02, 03:13-03:14:02, 03:16, 03:19:01-03:19:02, 03:21-03:22, 03:24, 03:27-03:29, 03:35-03:36, 03:42, 03:44, 03:46-03:60, 03:69, 03:71, 03:73, 03:75-03:77, 03:80, 03:82-03:84N, 03:92-03:94, 03:101-03:103, 03:108-03:109, 03:114-03:116, 03:118N-03:122, 03:127-03:131, 03:133-03:135, 03:138-03:140, 03:142-03:144, 03:147-03:148, 03:150, 03:152, 03:154, 03:157-03:160, 03:162-03:167, 03:169-03:173, 03:180, 03:182-03:183, 03:186-03:188, 03:191-03:198, 03:201-03:202, 03:206-03:208, 03:216, 03:218-03:219, 06:209
5 <sup>6</sup>	130 bp, 220 bp	<b>515 bp</b>	6, 8	*03:02:01-03:02:19, 03:05:01-03:05:04, 03:07-03:08, 03:11, 03:18, 03:32, 03:37, 03:45, 03:61, 03:63-03:64, 03:66N-03:68, 03:70, 03:85, 03:104, 03:106-03:107, 03:125, 03:132, 03:146, 03:153, 03:161, 03:174-03:175, 03:178-03:179, 03:181, 03:184-03:185, 03:189-03:190, 03:199, 03:203-03:205, 03:210-03:211, 03:213N-03:215, 03:220-03:221, 03:223-03:224, 03:226, 03:228-03:229, 06:29, 06:123, 06:139
6 <sup>6</sup>	135 bp	<b>515 bp</b>	2, 3, 6, 9	*02:03, 03:03:02:01-03:03:13, 03:06, 03:12, 03:15, 03:20, 03:25:01-03:26, 03:30-03:31, 03:33-03:34, 03:38-03:41, 03:43, 03:65, 03:74, 03:79, 03:86-03:91Q, 03:95N-03:99Q, 03:104-03:105, 03:111-03:113, 03:117, 03:123-03:124, 03:126, 03:136-03:137, 03:141, 03:145, 03:149, 03:155-03:156, 03:168, 03:176-03:177, 03:200, 03:209, 03:212, 03:222, 03:227, 03:230, 04:03:01-04:03:02, 06:03:10, 06:51:01, 06:66, 06:96, 06:168, 06:172

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<b>7<sup>5,6</sup></b>	90 bp	<b>515 bp</b>	3, 7, 8, 9	*03:01:01:01-03:01:06, 03:01:08-03:02:17, 03:02:19-03:05:04, 03:07, 03:09-03:24, 03:26-03:57, 03:59-03:64, 03:66N-03:103, 03:105-03:106, 03:108-03:136, 03:138-03:193, 03:195-03:227, 03:229-03:230
<b>8<sup>6,7</sup></b>	135 bp, 160 bp, 185 bp, 210 bp	430 bp	4	*03:132, 04:01:01-04:36N
<b>9</b>	225 bp	430 bp	4	*04:01:01-04:03:02, 04:06-04:21, 04:22 <sup>w</sup> , 04:23, 04:25N-04:32, 04:34-04:36N
<b>10</b>	215 bp	430 bp	5	*05:01:01:01-05:03:09, 05:03:11-05:03:17, 05:05:01-05:33, 05:35-05:43, 05:45-05:51, 05:53, 05:55-05:71, 05:73-05:76, 05:78-05:104, 05:106-05:113, 05:115-05:123, 06:156, 06:162, 06:169
<b>11</b>	185 bp	430 bp	8, 9	*03:03:11, 03:05:01, 03:05:03, 03:17:01, 03:61, 03:72, 03:100, 03:181
<b>12<sup>6</sup></b>	185 bp	430 bp	4, 7, 8, 9	*03:01:01:01-03:01:01:03, 03:01:03-03:01:07, 03:01:09-03:02:02, 03:02:04-03:02:12, 03:02:14-03:03:02:04, 03:03:04-03:04:02, 03:05:03-03:17:01, 03:18-03:19:02, 03:21-03:22, 03:23:02-03:36, 03:38-03:60, 03:62-03:71, 03:74, 03:76-03:98, 03:101-03:103, 03:106-03:108, 03:110-03:111, 03:113-03:117, 03:119-03:131, 03:133-03:153, 03:155, 03:157-03:161, 03:163-03:180, 03:182, 03:184-03:188, 03:190-03:203, 03:205-03:222, 03:224-03:225, 03:227-03:230, 04:01:03
<b>13</b>	185 bp	<b>515 bp</b>	6	*04:10, 06:02:01-06:02:25, 06:14:01-06:16, 06:19:01-06:20, 06:23-06:24, 06:33, 06:37, 06:46-06:50, 06:51:02, 06:68, 06:70-06:84, 06:95, 06:97, 06:107, 06:109, 06:111-06:117, 06:122, 06:124-06:127, 06:136-06:138, 06:146:01-06:147, 06:150-06:152, 06:156, 06:159, 06:161-06:163, 06:166, 06:173-06:175, 06:178-06:179N, 06:182-06:183, 06:188, 06:192, 06:197-06:198, 06:200-06:201, 06:208
<b>14</b>	185 bp	430 bp	1, 5, 6	*06:02:01-06:02:12, 06:02:14-06:03:10, 06:03:12-06:03:22, 06:05:02 <sup>?</sup> -06:06 <sup>?</sup> , 06:08:01-06:08:03, 06:10-06:11:03, 06:13:01-06:14:03, 06:16, 06:18:01-06:20, 06:23-06:24, 06:26N-06:27:02, 06:29-06:33, 06:37, 06:40-06:41, 06:44, 06:47-06:51:02, 06:59-06:65, 06:67-06:68, 06:70-06:78, 06:80-06:84, 06:87, 06:90-06:91, 06:95-06:97, 06:99:01-06:99:02, 06:106-06:107, 06:109-06:117, 06:122-06:128, 06:130-06:131, 06:133-06:134, 06:136-06:139, 06:141, 06:143, 06:145, 06:147-06:148, 06:150-06:152, 06:154, 06:156, 06:159, 06:161-06:163, 06:165-06:166, 06:169-06:170, 06:173-06:176, 06:178-06:179N,

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				06:182-06:185, 06:187-06:188, 06:190-06:192, 06:195-06:198, 06:200-06:201, 06:206, 06:208
<b>15</b>	135 bp	430 bp	3, 5, 6, 9	*03:10:01-03:10:02, 03:12, 03:14:01-03:14:02, 03:70, 03:179, 03:183, 03:195, 06:01:01-06:01:06, 06:01:08-06:01:15, 06:06 <sup>?</sup> , 06:43, 06:54N-06:58, 06:98, 06:99:02-06:105, 06:108, 06:120, 06:132, 06:140, 06:153, 06:157, 06:167-06:168, 06:177, 06:181, 06:194, 06:205, 06:209
<b>16<sup>8</sup></b>	-	-	-	<b>Negative Control</b>

<sup>1</sup>Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQ low resolution SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

<sup>2</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases. In the presence of a specific amplification the intensity of the control band often decreases.

<sup>3</sup>The serological split of the DQB1\*05:05 to 05:123, DQB1\*06:06 to 06:07, 06:10, 06:13, 06:15 to 06:24 and 06:27 to 06:209, the DQB1\*02:04 to 02:74 the DQB1\*03:02:02 to 03:02:04, 03:03:03, 03:05:02, 03:07 to 03:09 and 03:11 to 03:230 and the DQB1\*04:03:01 to 04:36N alleles is not known. The grouping of not serologically defined alleles is taken from the expert-assigned serological grouping in Tissue Antigens (2009) 73:95-170.

<sup>4</sup>For several DQB1 alleles 1<sup>st</sup> and/or 3<sup>rd</sup> exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

<sup>5</sup>HLA-Specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

<sup>6</sup>Primer mixes 5 to 8 and 12 may give a lower yield of HLA-specific PCR products than the other DQ low resolution primer mixes.

<sup>7</sup>The primer pairs in wells 1, 2 and 8 will in some samples give rise to two HLA-specific PCR fragments.

<sup>8</sup>Primer mix 16 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

'ser', serological HLA specificity

'w', might be weakly amplified.

'?', nucleotide sequence information not available for the primer matching sequence.



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### PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	135 230	135 185 220 270	210	130 220	130 220	135	90	135 160 185 210	225	215	185	185
Length of int. pos. control <sup>1</sup>	515	515	430	515	515	515	515	430	430	430	430	430
5'-primer(s) <sup>2</sup>	25(170) 5'-gCA 3'	9(122) 5'-gTT 3'	27(178) 5'-TgC 3'	26(173) 5'-TTA 3'	28(179) 5'-gAC 3'	26(173) 5'-TCT 3'	71(309) 5'-ACC 3'	21(159) 5'-ACC 3'	9(122) 5'-gTT 3'	29(184) 5'-gAC 3'	21(159) 5'-ACC 3'	38(210) 5'-gCA 3'
		26(173) 5'-ggA 3'	24(169) 5'-TgT 3'	29(184) 5'-gAg 3'		28(179) 5'-gAC 3'		71(309) 5'-ACC 3'	23(164) 5'-gCT 3'			
		26(173) 5'-ggg 3'	26(173) 5'-TCT 3'	30(185) 5'-AAg 3'		28(179) 5'-gAC 3'			38(210) 5'-gCg 3'			
			26(173) 5'-TTA 3'	30(185) 5'-AAA 3'								
3'-primer(s) <sup>3</sup>	57(266) 5'-CAA 3'	57(266) 5'-CAA 3'	86(353) 5'-gCT 3'	55(260) 5'-gCg 3'	57(266) 5'-Cgg 3'	57(266) 5'-CgT 3'	86(353) 5'-gCT 3'	69(304) 5'-CTC 3'	69(304) 5'-CTC 3'	87(356) 5'-ggA 3'	69(304) 5'-CCT 3'	86(353) 5'-gCT 3'
		87(356) 5'-ggT 3'	57(266) 5'-CAT 3'	86(353) 5'-gCT 3'	57(266) 5'-CAg 3'		86(354) 5'-AgT 3'	77(327) 5'-ACg 3'		87(356) 5'-ggT 3'		
		87(356) 5'-ggT 3'	86(353) 5'-ACC 3'	86(354) 5'-AgT 3'	57(266) 5'-Cgg 3'		86(355) 5'-gAC 3'					
		88(361) 5'-CCT 3'	86(353) 5'-ACg 3'		87(356) 5'-ggg 3'		87(356) 5'-ggC 3'					
			86(354) 5'-AAT 3'				87(358) 5'-gCC 3'					
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15
Length of spec. PCR product	185	185	135
Length of int. pos. control <sup>1</sup>	515	430	430
5'-primer(s) <sup>2</sup>	9(122) 5'-gTT 3'	38(209) 5'-CgC 3'	13(134) 5'-ggC 3'
3'-primer(s) <sup>3</sup>	57(266) 5'-CAT 3'	86(353) 5'-ACg 3'	45(230) 5'-CCC 3'
	58(270) 5'-TCC 3'		
Well No.	13	14	15

<sup>1</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

<sup>2</sup>The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>3</sup>The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

101.201-48/12 – including *Taq* pol., IFU-01  
 101.201-48u/12u – without *Taq* pol., IFU-02

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Lot No.: **4E0**

Lot-specific information

CELL LINE VALIDATION SHEET																			
DQ low resolution primer set <sup>2</sup>																			
				Well															
				1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	
				Production No.	201663501	201672402	201672403	201672404	201663505	201663506	201670407	201672408	201672409	201663510	201663511	201663512	201664713	201664714	201672415
IHC cell line <sup>1</sup>			DQB1																
1	9001	SA	*05:01		+	-	-	-	-	-	-	-	-	+	-	-	-	-	
2	9280	LK707	*06:01	*02:02	-	+	+	-	-	-	-	-	-	-	-	-	-	+	
3	9011	E4181324	*06:01		-	+	-	-	-	-	-	-	-	-	-	-	-	+	
4	9275	GU373	*02:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
5	9009	KAS011	*05:02		+	-	-	-	-	-	-	-	-	+	-	-	-	-	
6	9353	SM	*03:02	*06:01	-	+	-	-	+	-	+	-	-	-	-	+	-	+	
7	9020	QBL	*02:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
8	9025	DEU	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
9	9026	YAR	*03:02		-	-	-	-	+	-	+	-	-	-	-	+	-	-	
10	9107	LKT3	*04:01		-	-	-	-	-	-	-	+	+	-	-	-	-	-	
11	9051	PITOUT	*02:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
12	9052	DBB	*03:03		-	-	-	-	-	+	+	-	-	-	-	+	-	-	
13	9004	JESTHOM	*05:01		+	-	-	-	-	-	-	-	+	-	-	-	-	-	
14	9071	OLGA	*04:02		-	-	-	-	-	-	-	+	+	-	-	-	-	-	
15	9075	DKB	*03:03		-	-	-	-	-	+	+	-	-	-	-	+	-	-	
16	9037	SWEIG007	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
17	9282	CTM 3953540	*02:01	*06:03	-	+	+	-	-	-	-	-	-	-	-	-	+	-	
18	9257	32367	*06:02	*02:02	-	+	+	-	-	-	-	-	-	-	-	-	+	+	
19	9038	BM16	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
20	9059	SLE005	*06:04		-	+	-	-	-	-	-	-	-	-	-	-	-	-	
21	9064	AMALA	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
22	9056	KOSE	*05:03	*06:04	+	+	-	-	-	-	-	-	-	+	-	-	-	-	
23	9124	IHL	*05:03	*06:01	+	+	-	-	-	-	-	-	-	+	-	-	-	+	
24	9035	JBUSH	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
25	9049	IBW9	*02:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
26	9285	WT49	*02:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
27	9191	CH1007	*04:01	*05:01	+	-	-	-	-	-	-	+	+	+	-	-	-	-	
28	9320	BEL5GB	*02:02	*03:01	-	-	+	+	-	-	+	-	-	-	-	+	-	-	
29	9050	MOU	*02:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
30	9021	RSH	*04:02		-	-	-	-	-	-	-	+	+	-	-	-	-	-	
31	9019	DUCAF	*02:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
32	9297	HAG	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
33	9098	MT14B	*03:02		-	-	-	-	+	-	+	-	-	-	-	+	-	-	
34	9104	DHIF	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
35	9302	SSTO	*03:05		-	-	-	-	+	-	+	-	-	-	+	-	-	-	
36	9024	KT17	*03:02		-	-	-	-	+	-	+	-	-	-	-	+	-	-	
37	9065	HHKB	*06:03		-	+	-	-	-	-	-	-	-	-	-	-	+	-	
38	9099	LZL	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
39	9315	CML	*02:01	*03:01	-	-	+	+	-	-	+	-	-	-	-	+	-	-	
40	9134	WHONP199	*02:02	*03:03	-	-	+	-	-	+	+	-	-	-	-	+	-	-	
41	9055	H0301	*06:09		-	+	-	-	-	-	-	-	-	-	-	-	-	-	
42	9066	TAB089	*06:01		-	+	-	-	-	-	-	-	-	-	-	-	-	+	
43	9076	T7526	*03:03		-	-	-	-	-	+	+	-	-	-	-	+	-	-	
44	9057	TEM	*05:03		+	-	-	-	-	-	-	-	-	+	-	-	-	-	
45	9239	SHJO	*02:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	
46	9013	SCHU	*06:02		-	+	-	-	-	-	-	-	-	-	-	-	+	+	
47	9045	TUBO	*03:01		-	-	-	+	-	-	+	-	-	-	-	+	-	-	
48	9303	TER-ND	*05:01		+	-	-	-	-	-	-	-	-	+	-	-	-	-	

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**Lot No.: 4E0****Lot-specific information**

<sup>1</sup>The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

<sup>2</sup>The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

In primer solutions 1 to 3 and 5 one, two or three 5'-primers were not possible to test, and in primer solutions 1, 2, 4, 5, 7 and 13 one or more 3'-primers were not possible to test.

The specificities of the primers in primer solution 2 was tested by separately adding one additional 5'-primer, respectively one additional 3'-primer.

Additional 5'-primers in primer solution 7 was tested by separately adding another 3'-primer.

Additional 3'-primers in primer solutions 1 and 10 were tested by separately adding one 5'-primer.

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Lot No.: **4E0**

Lot-specific information

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